

VIEWPOINT: MULTIPLE-HARVEST SORGHUMS TOWARD IMPROVED FOOD SECURITY

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ABSTRACT

Perennial crops offer an avenue by which marginal lands might be brought into sustainable production while arresting or even reversing losses of ecological capital. The natural adaptation of sorghum to its native Sahelian region makes it essential to some of the poorest and most food-insecure

human populations, dependent upon some of the world's most degraded soils. Increased demand for limited fresh water, together with rising global temperature and aridity, suggest that the inherent drought tolerance of sorghum will be of growing importance. With some genotypes already known to be suitable for ratoon crop production, and with two sources of perenniality, the Sorghum genus has become a model for dissecting the molecular control of ratooning and perenniality. We are pursuing two broad approaches to investigate the efficacy of ratoon/perennial sorghum production and to develop genotypes suitable for these production systems. First, the identification of diploid progeny from crosses between S. bicolor and S. propinguum offers a starting point for investigation of these traits in conventional sorghums, by crossing to elite sorghums and evaluation of resulting progeny (potentially also employing DNA-marker aided selection to increase the frequency of genotypes that contain known Quantitative trait loci (QTLs) for ratooning and/or perenniality-related traits). Second, crosses between tetraploid forms of S. bicolor and their naturally-tetraploid relative S. halepense, may offer the possibility to benefit from many traits that differentiate this widespread weedy/invasive plant from its progenitors. While initial efforts have focused on grain production, substantial forage/biomass is also produced by ratoon/perennial sorghums and in principle there appears to be no insurmountable obstacle to also developing perennial 'sweet sorghums' suitable for sugar production. Perennial cropping systems may be especially attractive to smallholder agriculture (particularly benefitting women farmers), in which poor seedling emergence and stand establishment can lead to crop failure and costs of seed, energy, and equipment are limiting factors. However, the benefits of perennials may also apply to intensive agriculture, especially in situations where erosion, nutrient leaching, and declining organic matter levels occur. A fringe benefit of increased knowledge of perenniality may be new tools and approaches by which to control widespread agricultural weeds. Comparative data from sorghum and rice suggest that findings with regard to perenniality may extrapolate broadly to a wide range of taxa, accelerating progress in implementing new production systems (or weed control systems) in other genera.

Keywords: ratoon crop, perennial, Sorghum propinguum, Sorghum halepense

INTRODUCTION TO SORGHUM

About 20 million years ago, a population of 20-chromosome grassy plants had a singularly bright future. An early branch diverged from this population ~10-15 million years ago, experiencing genome-wide duplication and genome size expansion accompanied by chromosome number reduction leading to the modern maize genome (Gaut, Clark, et al. 1997; Swigonova, Lai, et al. 2004). About 8-9 million years ago (Jannoo et al. 2007; Sobral et al.



1994) another branch diverged, also experiencing genome-wide duplication and leading to the modern genomes of Saccharum. It was perhaps the most important biomass/biofuels crop worldwide, and its close relative *Miscanthus*, which is among the highest-yielding herbaceous temperate biomass crops known (Heaton, Dohleman, et al. 2008). A third branch spawned lineages with a wide range of fates (Spangler *et al.* 1999). One being domesticated into cultivated sorghum, and another experiencing polyploidy to form one of the world's most aggressive weeds (*Sorghum halepense*).

Among these important grasses, all members of the Andropogoneae clade, the genome of *Sorghum bicolor* L. Moench (sorghum) is of central importance both as a botanical model and as a major crop. Sorghum is the most drought-resistant of the world's top five cereal crops, and an important dual-purpose (grain + straw) crop. Its drought resistance is essential in the United States Southern Plains that often receive too little rain for other grains. And in arid countries of northeast and West Africa, sorghum contributes as much as 26-39 percent of calories in the human diet (www.fao.org). Nonetheless, sorghum improvement lags that of maize, wheat and rice, each of which have more than doubled in worldwide average yield in the last 38 years. Sorghum yields only gained 51 percent (www.fao.org). Increased demand for limited fresh water, together with rising global temperature and aridity, suggest that sorghum will be of growing importance.

While grain is of special importance, the value of other sorghum products should not be underestimated. Sweet sorghum for stock feed (indirectly sustaining protein production) is a large and growing market, and the possibility of production systems based on perenniality combined with low/no-till farming are very attractive. Further, bioenergy sorghums are a promising source of cellulosic ethanol (Rooney *et al.* 2007). They are even better suited to tropical regions (such as much of Africa) than temperate regions, because a more continuous supply of feedstock is possible in the tropics.

The sequenced ~740 Mb sorghum genome (Paterson *et al.* 2009) is a logical complement to that of *Oryza* (rice) for grass functional genomics. It is a representative of tropical grasses in which biochemical and morphological specializations ('C4' photosynthesis) improve carbon assimilation at high temperatures. The sorghum karyotype is suggested by parsimony to have been shared by the Andropogoneae common ancestor, with only ~3 percent differential sorghum-rice gene loss and less sorghum-rice structural rearrangement in 50 million years than sorghum-maize rearrangement in about 15 million years (Bowers *et al.* 2003). With 70 million years of 'abstinence' from genome duplication, functions of *S. bicolor* genes also may still resemble those of the common cereal ancestor.

Sorghum evolution and improvement have yielded a remarkable diversity of morphologies. Differences in inflorescence and seed morphology are used to classify *S. bicolor* races. Also they have been embellished by divergent selection applied to forms cultivated for purposes ranging from 'broomcorn' to the compact inflorescences and large seed of grain types. Divergent selection

has also affected plant stature, with stalks of up to 6 metres being a key building material in Africa, but less than 1 metre favoured for mechanized harvest. Selection for carbohydrate rich seed/grain has increased harvest index (carbon partition to grain) in grain types, while selection for sugar-rich stalks and/or total biomass has reduced seed size and output in others to an extreme exemplified by *S. propinquum* (Figure 1). Human selection has been superimposed on rich natural variation in plant architecture and carbon allocation, for example ranging from one to 100+ tillers per crown, and branching along a tiller ranging from none to secondary, tertiary, and even quaternary. Its morphological and biochemical diversity makes sorghum the only crop that is compatible with fuel production from each of three routes, including starch (seed), cellulose (stalks), and sugar ('sweet' sorghums have sugar yields comparable to those of sugarcane).

FIGURE 1. COMPARISON OF ANNUAL SORGHUM BICOLOR AND PERENNIAL S. PROPINQUUM

- (A) Crowns including extensive rhizomes of S. propinguum.
- (B) Seeds those of *S. bicolor* grain types are large and carbohydrate rich, while those of *S. propinquum* are very small.



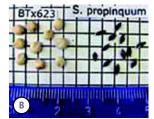
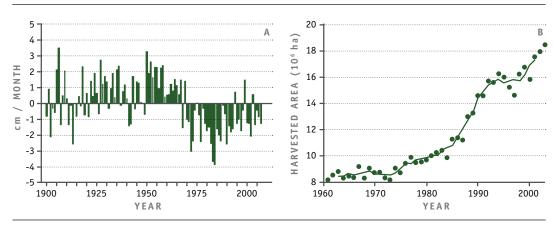


FIGURE 2. (A) SAHEL PRECIPITATION, 1900-2007; (B) SAHEL CROPPING AREA, 1960-2010



A: modified from Haywood et al., 2013 B: source: Kandji et al., 2006



Rich resources are in place to link sorghum phenotypic diversity to its molecular basis. More than 30 sorghum genomes and 40 transcriptomes, sampling all botanical races of cultivated sorghum, wild *S. bicolor*, and appropriate outgroups, have now been sequenced by the United States Department of Energy Joint Genome Institute. GBS has been performed on several reference collections (Morris *et al.* 2013), including two developed by the International Crops Research Institute for the Semi-Arid-Tropics (ICRISAT) that have been focal points of phenotyping for morphological variation, yield components, and drought responses (Upadhyaya *et al.* 2009).

OVERVIEW OF RESEARCH ON PERENNIALITY IN SORGHUM

The natural adaptation of sorghum to its native Sahelian region makes it essential to some of the poorest and most food-insecure human populations and dependent upon some of the world's most degraded soils. Years of above average rainfall were followed by drought in the Sahel starting in the late 1960s (Figure 2A). A dramatic increase in the Sahelian area devoted to crops began at that time, with falling yields necessitating greater effort to cultivate evermore fragile lands to feed growing populations (Figure 2B). These considerations are thought to contribute greatly to the fact that the Sahel now includes some of the most degraded lands on the planet.

Recently, the potential benefits of perennial grain crops have gained wider recognition (Glover et al. 2010). Perennial crops are essential to bringing marginal lands into sustainable production (Cox et al. 2002; Scheinost, 2001; Wagoner, 1990), maximizing ecosystem productivity (Field, 2001) and minimizing losses of topsoil (Pimentel et al. 1995), water, and nutrients. Perennial cover is >50 times more effective than annual crops in maintaining topsoil (Gantzer et al. 1990), and 30-50 times more effective than annuals at preventing nitrogen losses (Randall and Mulla, 2001). Perennial cropping systems (Cox et al. 2006; Jackson and Jackson, 1999; Piper, 1998) may be especially attractive to smallholder agriculture, in which poor seedling emergence and stand establishment can lead to crop failure, and costs of seed, energy, and equipment are limiting factors. It also may benefit mechanized agriculture in situations where erosion, nitrate (and other nutrient) leaching, and declining organic matter levels occur.

The *Sorghum* genus has become a model for dissecting the molecular control of ratooning and perenniality largely due to the availability of the diploid perennial *S. propinquum* which is abundantly tillering and has rhizomes that contribute to perenniality (Hu *et al.* 2003; Jang *et al.* 2006; Jang *et al.* 2008; Paterson *et al.* 1995). Tillers and rhizomes both develop from axillary buds at the lowermost nodes of the erect leafy shoot of the plant, with acropetally increasing tendency to develop into tillers (Gizmawy *et al.* 1985). There is appreciable correspondence in the locations of QTLs determining tillering and rhizomatousness (Paterson *et al.* 1995),

thought to be related to the number of buds available. The commitment of a bud to rhizome development, tiller development, or quiescence is genetically-determined (Paterson *et al.* 1995), and is associated with marked differences in gene expression (Jang *et al.* 2006). Finally, a high level of correspondence of rhizome QTLs between sorghum and rice (Hu *et al.* 2003), thought to have diverged from common ancestry about 50 million years ago, suggests that key genes conferring perenniality-related traits may be shared by many Poaceae cereals. In other words, fundamental information about these traits in one cereal may extrapolate broadly to many other cereals.

While no members of the cultivated species, *Sorghum bicolor*, are rhizomatous, there is variation among cultivars for ratoon crop productivity (Duncan *et al.* 1980; Duncan and Moss, 1987), and close relatives sugarcane and *Miscanthus* are usually grown as ratoon crops. Ratooning (regrowth) is a complex trait, showing quantitative inheritance (Duncan *et al.* 1980). Likewise, QTL mapping in an F2 population of a cross between an elite grain sorghum inbred BTx623 (later used for the reference sequence) and *S. propinquum*, showed regrowth to be closely related to both rhizomatousness and tillering, but was only able to account for 14-30 percent of phenotypic variance in these traits (Paterson *et al.* 1995). A recombinant inbred line population for this cross (Kong *et al.* 2013), now provides for replicated genetic analysis of productivity, and includes genotypes with a range of 'degrees' of ratoon cropping from near-zero to fully perennial. The recombinant inbred line population has been genetically mapped with both the population itself and map publicly available (Kong *et al.* 2013), and with three years of phenotyping for tillering, rhizomatousness, and basic metrics related to grain production (yield components) presently being analysed for peer-reviewed publication.

A relative of sorghum, *S. halepense*, is a wild perennial polyploid resulting from natural hybridization between *S. bicolor* (2n = 20) and *S. propinquum* (2n = 20), a wild perennial diploid native to Southeast Asia and estimated to have diverged from *S. bicolor* ~1-2 million years ago. *S. halepense* finds occasional use as forage and even food (seed/flour), but is most noted as one of the world's most noxious weeds, having spread from its west Asian centre of diversity across much of Asia, Africa, Europe, North and South America, and Australia. Its establishment in the United States of America is probably typical of its spread to other continents, including intentional introduction as a prospective forage and/or unintentional introduction as a contaminant of sorghum seedlots (McWhorter, 1971). However, while *S. bicolor* has remained largely confined to cultivation, *S. halepense* has readily naturalized – revealing genetic potential for adaptation that goes well beyond that of sorghum. *S. halepense* crosses readily with *S. bicolor* (Arriola and Ellstrand, 1996) –more than 90 percent of plants in naturalized *S. halepense* populations growing adjacent to long-term sorghum production fields contain alleles introgressed from sorghum (Morrell *et al.* 2005).

RESEARCH AGENDA

We are pursuing two broad approaches to investigate the efficacy of ratoon/perennial sorghum production, and to develop genotypes suitable for these production systems.

First, the identification of diploid progeny from crosses between *S. bicolor* and *S. propinquum* (Kong *et al.* 2013) offers a starting point for investigation of these traits in conventional sorghums, by crossing to elite sorghums and evaluation of resulting progeny (potentially also employing DNA-marker aided selection to increase the frequency of genotypes that contain known QTLs for ratooning and/or perenniality-related traits).

Second, crosses between tetraploid forms of *S. bicolor* and their naturally-tetraploid relative *S. halepense*, may offer the possibility to benefit from many traits that differentiate this widespread plant from its progenitors. While *S. halepense* is generally under selection for weediness-related attributes that are undesirable in annual row-crop production, some selection pressures may favour *S. halepense* alleles that are of potential value in sorghum, for example conferring rapid vegetative development and early flowering. An attractive but unproven hypothesis, which the ability of *S. halepense* to naturalize in a much wider range of environments than sorghum is cultivated may have been accompanied by selection for alleles associated with valuable attributes such as cold tolerance (for example in the northern United States of America and Russia), drought tolerance (Africa, Arabia, United States of America in the southwest), disease resistance (many) or other traits. Moreover, a host of data now show that unpredictable, presumably epistatic, effects of allele introgressions from seemingly undesirable sources can confer valuable traits to crops (Bernacchi *et al.* 1998a; Bernacchi *et al.* 1998b; Chee *et al.* 2005; Eshed and Zamir, 1995; Fridman *et al.* 2004; Fulton *et al.* 1997; Gur and Zamir, 2004; Schauer *et al.* 2006; Tanksley *et al.* 1996; Xiao *et al.* 1996; Zamir, 2001).

At present, we are phenotyping and analysing F2 populations, their F3 progeny and the progeny of backcrosses to *S. bicolor* for perenniality, plant architecture, yield components and domestication traits. While we find some of these tetraploids to have fecundity comparable to diploid sorghums and are potentially directly usable, another attractive route for their utilization would be to extract diploids with selected properties of *S. halepense* either by backcrossing or by the development of haploids.

IMPLICATIONS

The benefits of perenniality as a cropping system are receiving renewed interest. Perennial plants have four advantages over annuals: a longer growing season; better access to water and nutrients; more conservative use of nutrients and better adaptation to marginal lands (Cox *et al.* 2006). The first three of those characteristics are expressed on all landscapes, from prime farmland to poor or sloping soils unsuitable for annual cropping. The expansion of agriculture

to provide plant biomass for production of fuels or chemical feedstocks with little or no carbon release (Fargione *et al.* 2008) will require greater utilization of marginal land (Tilman *et al.* 2009), maximizing ecosystem productivity (Field, 2001) and minimizing losses of topsoil (Pimentel *et al.* 1995), water, and nutrients.

The benefits of perenniality apply to food, as well as biomass, crops. Annual food crops have dominated the agricultural landscape since the time of the earliest farmers. In the last few decades alone, one-third of the planet's arable land has been lost to soil erosion (Pimentel et al. 1995). No-till production of annual crops, designed to control soil loss, has so far required increased use of herbicides. Direct-seeded annual cropping systems produce as high or higher nitrate emissions as tillage systems (Mummey et al. 1998). 'Natural systems agriculture', based on developing a "domestic prairie" with herbaceous, perennial grain-producing crops grown in polyculture (Jackson and Jackson, 1999; Piper, 1998), has been proposed as a means to produce food on otherwise marginal lands while arresting or even reversing losses of ecological capital.

In smallholder production systems that are widespread in regions where sorghum is a staple in the human diet, conservation-agriculture approaches may particularly benefit women farmers, by reducing dependence on duties generally performed by men (ploughing) and mitigating the need for duties performed by women and children (uprooting grass, de-stumping) (Milder *et al.* 2011). Moreover, women frequently are given marginal fields that should be left fallow (Kevane, 2011), for example those which will be the greatest beneficiaries of multi-cropping approaches. Male farmers, with access to generally higher levels of land, labour, and cash, have a somewhat greater tendency to grow cash crops (Gilbert *et al.* 2005) – further indicating that early adopters of reduced-cost, reduced-labour multi-cropping sorghum production systems may tend to be female. A reduced requirement for seed selection in multi-cropping production systems (which propagate vegetatively), may also reduce male investment as seed selection is largely maledominated (Freeman and Boateng, 2012).

Perenniality plays two contrasting roles in agriculture, being: (i) a valuable asset in establishment and persistence of dense, productive stands of plants suitable for multiple harvests, and (ii) an essential component of "weediness" of many of our most noxious weeds. Better understanding of the biology of *S. halepense* itself, one of the most noxious weeds affecting world agriculture (Holm *et al.* 1977), may also be of high value. *S. halepense* is a major contaminant in sorghum seed production, an alternate host and means of over-wintering for pests and pathogens of both monocot and dicot crops. It is also a highly-effective competitor for sunlight and other resources, causing reductions in economic yield of 45 percent or greater in crops as diverse as monocots such as sugarcane (Millhollen, 1970) and dicots such as soybean (McWhorter and Hartwig, 1972). The first United States federal appropriation for weed control research targeted Johnsongrass (House Bill #121, 1900), yet it remains largely unchecked today.

The S. halepense problem constrains improvement of sorghum through biotechnology. S. halepense is a particular problem in and around sorghum fields because no known herbicide



can selectively eliminate it without damaging the sorghum. Transformation of sorghum was first accomplished in 1993 (Casas *et al.* 1993), and introduction of transgenes for resistance to insects, diseases, or herbicides could have a major impact on sorghum productivity. However, the risk of transgene escape constrains the ability of regulatory agencies to approve the release of sorghum cultivars with transgenes that could enhance agricultural productivity.

Better understanding of the biology of *S. halepense* and associated new functional genomics tools might aid greatly in identification of genes, regulatory elements, and biochemical functions that are important to its weediness, particularly its rhizomes, but dispensable to other plant processes (Jang *et al.* 2006). Identification of such genes would provide the foundation to search for plant growth regulators that specifically target rhizomes, perhaps providing for control of rhizomatous weeds even in closely related crops such as *S. halepense* in sorghum. Moreover, better understanding of rhizome development may also benefit development of plant genotypes optimal for biomass/biofeedstock production on marginal lands, as well as improvement of a host of forage and turf grasses.

Our demonstration that most QTLs responsible for rhizomatousness in *Sorghum* and *Oryza* respectively, map to corresponding locations (Hu *et al.* 2003), suggests that *information about rhizomatousness from a few models (that are also major crops) may extrapolate broadly to a wide range of taxa*. Successes in clarifying the genetics of perenniality in this genus, may accelerate progress in implementing new production systems in other genera, and also to controlling a wide range of related agricultural weeds.

REFERENCES

- Arriola, P.E. & Ellstrand, N.C. 1996. Crop-to-weed gene flow in the genus Sorghum (Poaceae): Spontaneous interspecific hybridization between johnsongrass, Sorghum halepense, and crop sorghum, S. bicolor. American Journal of Botany. 83: 1153-1159.
- Bernacchi, D., Beck-Bunn, T., Emmatty, D., Eshed, Y., Inai, S., Lopez, J. Petiard, V., Sayama, H., Uhlig, J., Zamir, D., & Tanksley, S. 1998. Advanced back-cross QTL analysis of tomato. II. Evaluation of near-isogenic lines carrying single-donor introgressions for desirable wild QTL-alleles derived from Lycopersicon hirsutum and L-pimpinellifolium (vol 97, pg 170, 1998). *Theoretical and Applied Genetics*. 97: 1191-1196.
- Bernacchi, D., Beck-Bunn, T., Eshed, Y., Lopez, J., Petiard, V., Uhlig, J., Zamir, D. & Tanksley, S.D. 1998a. Advanced backcross QTL analysis in tomato. I: Identification of QTLs for traits of agronomic importance from Lycopersicon hirsutum. *Theoretical and Applied Genetics*. 97:381-397.
- Bernacchi, D., Beck-Bunn, T., Emmatty, D., Eshed, Y., Inai, S., Lopez, J., Petiard, V., Sayama, H., Uhlig, J., Zamir, D. & Tanksley, S.D. 1998b. Advanced backcross QTL analysis in tomato. II: Evaluation of near-isogenic lines carrying single-donor introgressions for desirable wild QTL-alleles derived from Lycopersicon hirsutum and L. pimpinellifolium. *Theoretical and Applied Genetics*. 97: 170-180.
- Bowers, J.E., Abbey, C., Anderson, S., Chang, C., Draye, X., Hoppe, A.H., Jessup, R., Lemke, C., Lennington, J., Li, Z.K., Lin, Y.R., Liu, S.C., Luo, L.J., Marler, B.S., Ming, R.G., Mitchell, S.E., Qiang, D., Reischmann, K., Schulze, S.R., Skinner, D.N., Wang, Y.W., Kresovich, S., Schertz, K.F. & Paterson, A.H. 2003. A high-density genetic recombination map of sequence-tagged sites for Sorghum, as a framework for comparative structural and evolutionary genomics of tropical grains and grasses. Genetics 165: 367-386.
- Casas, A.M., Kononowicz, A.K., Zehr, U.B., Tomes, D.T., Axtell, J.D., Butler, L.G., Bressan, R.A. & Hasegawa, P.M. 1993. Transgenic sorghum plants via microprojectile bombardment. *Proceedings of the National Academy of Sciences of the United States of America*. 90(23):11212–11216.
- Chee, P., Draye, X., Jiang, C.X., Decanini, L., Delmonte, T.A., Bredhauer, R., Smith, C.W. & Paterson, A.H. 2005. Molecular dissection of interspecific variation between Gossypium hirsutum and Gossypium barbadense (cotton) by a backcross-self approach: I. Fiber elongation. *Theoretical and Applied Genetics*. 111: 757–763.
- Chee, P.W., Draye, X., Jiang, C.X., Decanini, L., Delmonte, T.A., Bredhauer, R., Smith, C.W. & Paterson, A.H. 2005. Molecular dissection of phenotypic variation between Gossypium hirsutum and Gossypium barbadense (cotton) by a backcross-self approach: III. Fiber length. *Theoretical and Applied Genetics*111: 772–781.
- Cox, T.S., Bender, M., Picone, C., Van Tassel, D.L., Holland, J.B., Brummer, E.C., Zoeller, B.E., Paterson, A.H. & Jackson, W. 2002. Breeding perennial grain crops. *Critical Reviews in Plant Sciences*. 21: 59-91.
- Cox, T.S., Glover, J.D., Van Tassel, D.L., Cox, C.M. & DeHaan, L.R. 2006. Prospects for developing perennial grain crops. *BioScience*. 56: 649-659.
- Draye, X., Chee, P., Jiang, C.X., Decanini, L., Delmonte, T.A., Bredhauer, R., Smith, C.W. & Paterson, A.H. 2005. Molecular dissection of interspecific variation between Gossypium hirsutum and G. barbadense (cotton) by a backcross-self approach: II. Fiber fineness. *Theoretical and Applied Genetics*. 111: 764–771.
- **Duncan, R.R., Miller, F.R. & Bockholt, A.J.** 1980. Inheritance of tiller regrowth in ratooned sorghum. *Canadian Journal of Plant Science*. 60: 473-478.
- Duncan, R.R., & Moss, R.B. 1987. Comparative yields of ration cropped temperately and tropically adapted grain-sorghum hybrids. Crop Science. 27: 569-571.
- Eshed, Y. & Zamir, D. 1995. An introgression line population of lycopersicon pennellii in the cultivated tomato enables the identification and fine mapping of yield-associated qtl. *Genetics*. 141: 1147-1162.



- Fargione, J., Hill, J., Tilman, D., Polasky, D. & Hawthorne, P. 2008. Land clearing and the biofuel carbon debt. Science. 319: 1235-1238.
- Field, C.B. 2001. Sharing the garden. Science. 294: 2490-2491.
- Freeman, C.K. & Boateng, S.D. 2012. Gender and cultural dimensions of sorghum seed management in north-east Ghana. Global Journal of Biology, Agriculture and Health Sciences. 1: 4-9.
- Fridman, E., Carrari, F., Liu, Y.S., Fernie, A.R. & Zamir, D. 2004. Zooming in on a quantitative trait for tomato yield using interspecific introgressions. Science. 305: 1786-1789.
- Fulton, T.M., BeckBunn, T., Emmatty, D., Eshed, Y., Lopez, J., Petiard, V., Uhlig, J., Zamir, D. & Tanksley, S.D. 1997. QTL analysis of an advanced backcross of Lycopersicon peruvianum to the cultivated tomato and comparisons with QTLs found in other wild species. *Theoretical and Applied Genetics*. 95: 881-894.
- Gantzer, C.J., Anderson, S.H., Thompson, A.L. & Brown, J.R. 1990. Estimating soil erosion after 100 years of cropping on Sanborn Field. *Journal of Soil and Water Conservation*. 45: 641-644.
- Gaut, B.S., Clark, L.G., Wendel, J.F. & Muse, S.V. 1997. Comparisons of the molecular evolutionary process at rbcL and ndhF in the grass family (Poaceae). *Molecular Biology and Evolution*. 14: 769-777.
- Gilbert, R., Sakala, W.D. & Benson, T.D. 2005. Gender analysis of a nationwide cropping system trial survey in Malawi. *African Studies Quarterly*. 6: web.africa.ufl.edu/asq/v6/v6i1a9.htm
- **Gizmawy, I., Kigel, J., Koller, D. & Ofir, M.** 1985. Initiation, orientation, and early development of primary rhizomes in *Sorghum halepense* (L.) *Annals of Botany*. 55: 343.
- Glover, J.D., Reganold, J.P., Bell, L.W., Borevitz, J., Brummer, E.C., Buckler, E.S., Cox, C.M., Cox, T.S., Crews, T.E., Culman, S.W., DeHaan, L.R., Eriksson, D., Gill, B.S., Holland, J., Hu, F., Hulke, B.S., Ibrahim, A.M.H., Jackson, W., Jones, S.S., Murray, S.C., Paterson, A.H., Ploschuk, E., Sacks, E.J., Snapp, S., Tao, D., Van Tassel, D.L., Wade, L.J., Wyse, D.L. & Xu, Y. 2010. Increased food and ecosystem security via perennial grains. *Science*. 328: 1638-1639.
- Gur, A. & Zamir, D. 2004. Unused natural variation can lift yield barriers in plant breeding. Plos Biology. 2: 1610-1615.
- Hash, C.T., Ramu, P., Folkertsma, R.T., Upadhyaya, H.D., Billot, C., Rami, J-F., Deu, M., Gardes, L., Rivallan, R., Li, Y., Wang, T. & Lu, P. 2008. Diversity analysis of the sorghum global composite collection and reference set. 2008 Annual Research Meeting Generation Challege Programme. Bangkok, Thailand.
- Haywood, J.M., Jones, A., Bellouin, N. & Stephenson, D. 2013. Asymmetric forcing from stratospheric aerosols impacts Sahelian rainfall. *Nature Climate Change*. 3: 660-665. doi:10.1038/nclimate1857.
- **Heaton, E.A., Dohleman, F.G. Long, S.P.** 2008. Meeting US biofuel goals with less land: the potential of Miscanthus. *Global Change Biology*. 14: 2000-2014.
- Holm, L.G., Plucknett, D.L., Pancho, J.V. & Herberger, J.P. 1977. The world's worst weeds: distribution and biology. *University Press of Hawaii*. Honolulu, HI.
- Hu, F.Y., Tao, D.Y. Sacks, E. Fu, B.Y. Xu, P., Li, J., Yang, Y., McNally, K., Khush, G.S., Paterson, A.H. & Li, Z-K. 2003. Convergent evolution of perenniality in rice and sorghum. Proceedings of the National Academy of Sciences of the United States of America. 100: 4050-4054.
- Jackson, W. & Jackson, L.L. 1999. Developing high seed yielding perennial polycultures as a mimic of mid-grass prairie. In: Lefroy, E.C., Hobbs, R J., O'Connor, M. H., & Pate, J. S., editors, Agriculture as a Mimic of Natural Systems. Kluwer Academic Publishers, Dordrecht, Netherlands. pp. 1-37.
- Jang, C.S., Kamps, T.L Skinner, D.N., Schulze, S.R., Vencill, W. & Paterson, A.H. 2006. Sorghum genes with rhizome-enriched expression: functional classification, genomic organizatin, putative cis-acting regulatory elements, and relationship to QTLs. *Plant Physiology*. 142: 1148-1159.
- Jang, C.S., Kamps, T.L., Tang, H., Bowers, J.E., Lemke, C. & Paterson, A.H. 2008. Evolutionary fate of rhizome-specific genes in a non-rhizomatous Sorghum genotype. *Heredity*. 102: 266-273.

- Jannoo, N., Grivet, L., Chantret, N., Garsmeur, O., Glaszmann, J.C., Arruda, P. & D'Hont, A. 2007. Orthologous comparison in a gene-rich region among grasses reveals stability in the sugarcane polyploid genome. *The Plant Journal*. 50: 574-585.
- Kandji, S.T., Verchot, L. & Mackensen, J. 2006. Climate Change and Variability in the Sahel Region: Impacts and Adaptation Strategies in the Agricultural Sector. United Nations Environment Program (UNEP), and World Agroforestry Center (ICRAF), Nairobi, Kenya. p. 47.
- **Kevane, M.** 2011. Gendered production and consumption in rural Africa. *Proceedings of the National Academy of Sciences of the United States of America*. 109: 12350-12355. doi: 10.1073/pnas.1003162108.
- Kong, W., Jin, H., Franks, C.D., Kim, C., Bandopadhyay, R., Rana, M.K., Auckland, S.A., Goff, V.H., Rainville, L.K., Burow, G.B., Woodfin, C., Burke, J.J. & Paterson, A.H. 2013. Genetic analysis of recombinant inbred lines for *Sorghum bicolor* x *S. propinguum*. *G3*. 3: 101-108.
- McWhorter, C.G. 1971. Introduction and spread of Johnsongrass in the United States. Weed Science. 19: 496.
- **McWhorter, C.G. & Hartwig, E.E.** 1972. Competition of johnsongrass and cocklebur with six soybean varieties. *Weed Science.* 20: 56-59.
- Milder, J.C., Majanen, T. & Scherr, S.J. 2011. Performance and potential of conservation agriculture for climate change adaptation and mitigation in sub-Saharan Africa. An assessment of WWF and CARE projects in support of the WWF-CARE Alliance's Rural Futurres Initiative. (Available at https://foodgrainsbank.merchantsecure.com/uploads/CARE-WWF-EcoAgriculture_Conservation_Agriculture_in_Sub-Saharan_Africa_FINAL_REPORT_2011- 02-2811.pdf). p. 90.
- Millhollen, R.W. 1970. MSMA for johnsongrass control in sugarcane. Weed Science. 18: 333.
- Morrell, P.L., Williams-Coplin, D., Bowers, J.E., Chandler, J.M. & Paterson, A.H. 2005. Crop-to-weed introgression has impacted allelic composition of johnsongrass populations with and without recent exposure to cultivated sorghum. *Molecular Ecology*. 14: 2143-2154.
- Morris, G.P., Ramu, P., Deshpande, S.P., Hash, C.T., Shah, T., Upadhyaya, H.D., Lizarazu, O., Brown, P.J., Acharya, C.B., Mitchell, S.E., Harriman, J., Glaubitz, J.C., Buckler, E.S. & Kresovich, S. 2013. Population genomic and genome-wide association studies of agroclimatic traits in sorghum. *Proceedings of the National Academy of Sciences of the United States of America*. 110: 453-458.
- Mummey, D.L., Smith, J.L. & Bluhm, G. 1998. Assessment of alternative soil management practices on N₀ emissions from US agriculture. *Agriculture Ecosystems & Environment*. 70: 79-87.
- Paterson, A.H., Bowers, J.E., Bruggmann, R., Dubchak, I., Grimwood, J., Gundlach, H., Haberer, G., Hellsten, U., Mitros, T., Poliakov, A., Schmutz, J., Spannagl, M., Tang, H., Wang, X., Wicker, T., Bharti, A.K., Chapman, J., Feltus, F.A., Gowik, U., Grigoriev, I.V., Lyons, E., Maher, C.A., Martis, M., Narechania, A., Otillar, R.P., Penning, B.W., Salamov, A.A., Wang, Y., Zhang, L., Carpita, N.C., Freeling, M., Gingle, A.R., Hash, C.T., Keller, B., Klein, P., Kresovich, S., McCann, M.C., Ming, R., Peterson, D.G., Mehboob-ur-Rahman, Ware, D., Westhoff, P., Mayer, K.F., Messing, J. & Rokhsar, D.S. 2009. The Sorghum bicolor genome and the diversification of grasses. Nature. 457: 551-556.
- Paterson, A.H., Schertz, K.F., Lin, Y.R., Liu, S.C. & Chang, Y.L. 1995. The weediness of wild plants molecular analysis of genes influencing dispersal and persistence of Johnsongrass, Sorghum halepense (L) Pers. In Proceedings of the National Academy of Sciences of the United States of America. 92: 6127-6131.
- Pimentel, D., Harvey, C., Resosudarmo, P., Sinclair, K., Kurz, D., McNair, M., Crist, S., Shpritz, L., Fitton, L., Saffouri, R. & Blair, R. 1995. Environmental and economic costs of soil erosion and conservation benefits. Science. 267: 1117-1123.
- **Piper, J.K.** 1998. Growth and seed yield of three perennial grains within monocultures and mixed stands. *Agriculture Ecosystems & Environment*. 68: 1-11.
- Randall, G.W. & Mulla, D. 2001. Nitrate nitrogen in surface waters as influenced by climatic conditions and agricultural practices. *Journal of Environmental Quality*. 30: 337-344.
- Rooney, W.L., Blumenthal, J., Bean, B. & Mullet, J.E. 2007. Designing sorghum as a dedicated bioenergy feedstock. *Biofuels Bioproducts & Biorefining-Biofpr.* 1: 147-157. doi:10.1002/bbb.15.



- Schauer, N., Semel, Y., Roessner, U., Gur, A., Balbo, I., Carrari, R., Pleban, T., Perez-Melis, A., Bruedigam, C., Kopka, J., Willmitzer, L., Zamir, D. & Fernie, A.R. 2006. Comprehensive metabolic profiling and phenotyping of interspecific introgression lines for tomato improvement. *Nature Biotechnology*. 24: 447-454.
- Scheinost, P. 2001. Perennial wheat: a sustainable cropping system for the Pacific Northwest. *American Journal of Alternative Agriculture*. 16: 147-151.
- Sobral, B.W.S., Braga, D.P.V., Lahood, E.S. & Keim, P. 1994. Phylogenetic analysis of chloroplast restriction enzyme site mutations in the Saccharinae Griseb subtribe of the Andropogoneae Dumort tribe. *Theoretical and Applied Genetics.* 87: 843-853.
- Spangler, R., Zaitchik, B., Russo, E. & Kellogg, E. 1999. Andropogoneae evolution and generic limits in Sorghum (Poaceae) using ndhF sequences. *Systematic Botany*. 24: 267-281.
- Swigonova, Z., Lai, J., Ma, J., Ramakrishna, W., Llaca, V., Bennetzen, J.L. & Messing, J. 2004. Close split of sorghum and maize genome progenitors. *Genome Research*. 14: 1916-1923.
- Tanksley, S.D., Grandillo, S., Fulton, T.M., Zamir, D., Eshed, Y., Petiard, V., Lopez, J. & Beck-Bunn, T. 1996. Advanced backcross QTL analysis in a cross between an elite processing line of tomato and its wild relative L-pimpinellifolium. *Theoretical and Applied Genetics*. 92: 213-224.
- Tilman, D., Socolow, R., Foley, J.A., Hill, J., Larson, E., Lynd, L., Pacala, S., Reilly, J., Searchinger, T., Somerville, C., & Williams, R. 2009. Beneficial biofuels the food, energy, and environment trilemma. Science. 325: 270-271.
- Upadhyaya, H.D., Pundir, R.P.S., Dwivedi, S.L., Gowda, C.L.L., Reddy, V.G. & Singh, S. 2009. Developing a mini core collection of sorghum for diversified utilization of germplasm. Crop Science. 49: 1769-1780.
- **Wagoner, P.** 1990. Perennial grain development Past efforts and potential for the future. *Critical Reviews in Plant Sciences*. 9: 381-408.
- Xiao, J., Li, J., Grandillo, S., Ahn, S.N., McCouch, S., Tanksley, S. & Yuan, L.P. 1996. A wild species contains genes that may significantly increase the yield of rice. *Nature*. 384: 223-224.
- Zamir, D. 2001. Improving plant breeding with exotic genetic libraries. Nature Reviews Genetics. 2: 983-989.